



Instructions For Use APS-2-IFU-IVD

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Alcian Blue – PAS Stain Kit

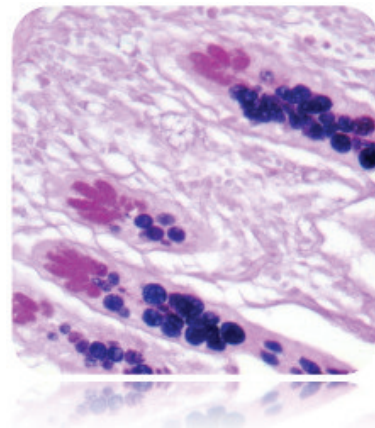
Description:

The Alcian Blue - PAS Stain Kit is intended for use in the simultaneous histological visualization of sulfated and carboxylated acid mucopolysaccharides, sulfated and carboxylated sialomucins (glycoproteins) and neutral mucins.

Acidic Sulfated Mucosubstances:	Blue
Hyaluronic Acid:	Blue
Sialomucins:	Blue
Neutral Mucins:	Magenta
Mixtures of Acidic and Neutral Mucins:	Blue - Mauve depending on dominant entity.

Uses/Limitations:

Not to be taken internally.
For In-Vitro Diagnostic use only.
Histological applications.
Do not use if reagents become cloudy.
Do not use past expiration date.
Use caution when handling reagents.
Non-Sterile.



Control Tissue:

Small or large Intestine
Appendix
Colon

Ordering information regarding individual components on back page!

Kit Contents:

<u>Item #</u>	<u>Kit Contents</u>	<u>Volume</u>	<u>Storage</u>
AAG030	Acetic Acid Solution (3%)	30 ml	18-25 °C
ANC030	Alcian Blue Solution, pH 2.5	30 ml	18-25 °C
PAQ030	Periodic Acid Solution (1%)	30 ml	2-8 °C
SRF030	Schiff's Solution	30 ml	2-8 °C
HMM030	Hematoxylin, Mayer's (Lillie's)	30 ml	18-25 °C

Precautions:

Avoid contact with skin and eyes.
Harmful if swallowed.
Follow all Federal, State, and local regulations regarding disposal.

Storage: 2° C



25° C

**Mixed Storage Conditions.
Separate Contents.**

Doc: IFU-TemplateMixedStoragerev2



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Procedure (Room Temperature):

1. Deparaffinize sections if necessary and hydrate to distilled water.
 2. Apply 5-10 drops of Acetic Acid Solution (3%) to tissue section for 2 minutes.
 3. Remove excess Acetic Acid Solution (3%) and without rinsing apply 5-10 drops of Alcian Blue (pH 2.5) solution for 15-20 minutes.
 4. Rinse for 2 minutes in running tap water followed by 2 changes of distilled water.
 5. Apply 5-10 drops of Periodic Acid Solution to tissue section for 5 minutes.
 6. Rinse slide in 2 changes of distilled water.
 7. Apply 5-10 drops of Schiff's Solution to tissue section for 10-20 minutes.
 8. Rinse for 2 minutes in warm running tap water followed by 2 changes of distilled water.
- Note: In some cases small crystals will form on the glass surface resulting in the appearance of a slight "film" which can be easily removed with 1-2 quick dips in a 1% Sulfuric Acid in water solution followed by a rinse in distilled water.
9. Apply 5-10 drops of Hematoxylin, Mayer's (Lillie's Modification) to tissue section for 2 minutes.
 10. Rinse for 2 minutes in running tap water followed by 2 changes of distilled water.
 11. Dehydrate through graded alcohols.
 12. Clear, and mount in synthetic resin.

References:

1. Lillie, R.D. 1977, H.J. Conn's Biological Stains, 9th Edition. Williams & Wilkins, Baltimore. Pages 452-455.
2. Sheenan, D.C., Hrapchak, B.B. Theory and Practice of Histotechnology, 2nd Edition. Battelle Press, Columbus, OH. Pages 172-173.
3. Churukian, C.J., 1989, Manual of Special Stains Laboratory, 4th Edition. University of Rochester, Rochester, New York. Pages 55-56.
4. Carson, F.L., 1996, Histotechnology; A Self-Instructional Text, 2nd Edition. ASCP Press, Chicago, IL. Pages 117-121.

Bulk Reagent Ordering Information and Current Pricing at www.scytek.com

Description:	Catalog #	Volume
Acetic Acid Solution (3%)	AAG500	500 ml
	AAG999	1000 ml
Alcian Blue Solution, pH 2.5	ANC250	250 ml
	ANC500	500 ml
	ANC999	1000 ml
	ANC3800	1 Gallon
Periodic Acid Solution (1%)	PAQ250	250 ml
	PAQ500	500 ml
	PAQ999	1000 ml
Schiff's Solution	SRF250	250 ml
	SRF500	500 ml
	SRF999	1000 ml
Hematoxylin, Mayer's (Lillie's Modification)	HMM500	500 ml
	HMM999	1000 ml
	HMM3800	1 Gallon

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25° C

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