P.O. Box 3286 - Logan, Utah 84323, U.S.A. - Tel. (800) 729-8350 – Tel. (435) 755-9848 - Fax (435) 755-0015 - www.scytek.com

Copper Stain Kit (For Microwave)

Description: The Copper Stain Kit (For Microwave) is intended for the demonstration of copper deposits in tissue sections.

Copper Deposits:	Light Brown to Red
Nuclei:	Blue

Uses/Limitations:
Not to be taken internally.
For In-Vitro Diagnostic use only.
Histological applications.
Do not use past expiration date.
Use caution when handling reagents.
Non-Sterile

Control Tissue: Fetal Liver or a known positive.

Availability/Contents:

Item #	Kit Contents	Volume	Storage
RSS030	Rhodanine Solution (Stock)	30 ml	2-8°C
SAB500	Acetate Buffer Solution, pH 8.0	2 x 500 ml	18-25°C
HMM125	Hematoxylin, Mayer's (Lillie's Mod.)	125 ml	18-25°C

Precautions:
Keep away from open flame.
Avoid contact with skin and eyes.
Harmful if swallowed.
Follow all Federal, State, and local regulations regarding disposal.
Use in chemical fume hood whenever possible.

Procedure (Standard):

Prepare Working Rhodanine Solution:

Combine:

4 ml Rhodanine Solution (Stock). Shake Stock Solution immediately before adding to Acetate Buffer.

46 ml Acetate Buffer Solution, pH 8.0

1. Deparaffinize sections if necessary and hydrate to distilled water.
2. Place loosely capped staining jar containing Working Rhodanine in microwave and heat solution until warm but not hot.
3. Place slide in warmed Working Rhodanine Solution and microwave at full power until solution is hot. Do not allow solution to boil.
4. Cap container, gently agitate to mix evenly, and allow solution to cool on countertop to room temperature with occasional agitation.
5. Examine slide microscopically and repeat heating/cooling cycle (steps 3 & 4) until desired staining intensity is achieved.

ScyTek Laboratories, Inc.
205 South 600 West
Logan, UT 84321
U.S.A.

CE IVD
EC REP EmergoEurope (31)(0) 70 345-8570
Molsnstraat 15
2513 BH Hague, The Netherlands

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6. Rinse slide in 2 changes of Acetate Buffer Solution, pH 8.0 for 1 minute each.
7. Stain tissue section with Hematoxylin, Mayer's (Lillie's Modification) for 5-10 seconds.
8. Rinse slide in 3 changes of Acetate Buffer Solution, pH 8.0 for 1 minute each.
9. Dehydrate slide in 3 changes of absolute alcohol.
10. Clear in 2 changes of xylene or xylene substitute, and mount in synthetic resin.

References:

1. Sheehan, DC., Hrapchak, BB. Theory and Practice of Histotechnology; 1980, page 230.
2. Lindquist, RR. Studies on the Pathogenesis of Hepatolenticular II: Cytochemical methods for the location of copper. Arch Pathol; 1969, Volume 87: page 370.

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