P.O. Box 3286 - Logan, Utah 84323, U.S.A. - Tel. (800) 729-8350 – Tel. (435) 755-9848 - Fax (435) 755-0015 - www.scytek.com

Giemsa Stain Kit (May-Grunwald)

(For Bone Marrow)

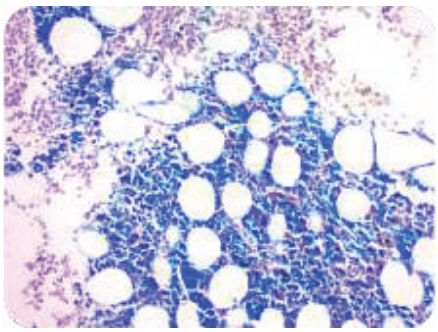
Description:

The Giemsa Stain Kit (May-Grunwald) is intended for use in the visualization of cells present in hematopoietic tissues and certain microorganisms. This kit may be used on formalin-fixed, paraffin-embedded or frozen sections.

Nuclei:	*Blue/Violet*
Cytoplasm	*Light Blue*
Collagen:	*Pale Pink*
Muscle Fibers:	*Pale Pink*
Erythrocytes:	*Gray, Yellow or Pink*
Rickettsia:	*Reddish-Purple*
Helicobacter Pylori:	*Blue*
Mast Cells:	*Dark Blue with Red Granules*

Uses/Limitations:

Not to be taken internally
For In-Vitro Diagnostic use only.
Histological applications.
Do not use past expiration date.
Use caution when handling these reagents.
Do not use if reagents become cloudy.
Non-Sterile



Control Tissue:

Blood Film
Bone Marrow
Spleen
Any well fixed tissue.

Availability/Contents:

Item #	Kit Contents	Volume	Storage
MAY030	*May-Grunwald Stock Solution*	*30 ml*	*18-25°C*
GGS008	*Giemsa Stock Solution*	*8 ml*	*18-25°C*
PBM060	*Phosphate Buffer Solution, pH 6.8*	*2 x 60 ml*	*18-25°C*
	Graduated Mixing Vial	*2*	

Precautions:

Keep away from open flame.
Avoid contact with skin and eyes.
Harmful if swallowed.
Follow all Federal, State, and local regulations regarding disposal.
Use in chemical fume hood whenever possible.

Storage: 18° C 25° C

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Preparation of Reagents Prior to Beginning:

1. *Prepare* **Working May-Grunwald Solution** *by mixing 10 drops of May-Grunwald Solution (MAY030) with 10 drops of Phosphate Buffer Solution, pH 6.8 (PBM060). Mix by agitating.*

2. *Prepare* **Working Giemsa Solution** *by mixing 1 drop of Giemsa Stock Solution (GGS030) with 20 drops of Phosphate Buffer Solution, pH 6.8 (PBM060). Mix by agitating.* **If staining a peripheral blood smear**, *instead use 3 drops of Giemsa Stock Solution (GGS030) with 20 drops of Phosphate Buffer Solution, pH 6.8 (PBM060).*

Procedure (Standard):

1. *Deparaffinize sections if necessary and hydrate to distilled water.*
2. *Place slide in staining tray and flood with Working May-Grunwald Solution for 6 minutes. Note: Agitate slide occasionally to insure proper staining.*
3. *Flood slide with Phosphate Buffer Solution, pH 6.8 until no stain runs off.*
4. *Flood slide with Working Giemsa Solution for 13 minutes. Note: Agitate slide occasionally to insure proper staining.*
5. *Flood slide with Phosphate Buffer Solution, pH 6.8 until no stain runs off.*
6. *Allow Phosphate Buffer Solution, pH 6.8 to remain on slide for an additional 3 minutes.*
7. *Dip slide quickly in distilled water and air dry at room temperature.*
8. *Dip slide in Xylene or Xylene Substitute.*
9. *Mount in synthetic resin.*

Procedure (Mast Cells):

1. *Deparaffinize sections if necessary and hydrate to distilled water.*
2. *Place slide in staining tray and flood with Working May-Grunwald Solution for 6 minutes. Note: Agitate slide occasionally to insure proper staining.*
3. *Flood slide with Phosphate Buffer Solution, pH 6.8 until no stain runs off.*
4. *Flood slide with Working Giemsa Solution for 13 minutes. Note: Agitate slide occasionally to insure proper staining.*
5. *Flood slide with Phosphate Buffer Solution, pH 6.8 until no stain runs off.*
6. *Differentiate by dipping slide in Acetic Acid Solution (0.25%) until background is desired intensity.*
7. *Flood slide with Phosphate Buffer Solution, pH 6.8 for 10-15 seconds.*

Storage: 18° C – 25° C

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8. *Dip slide quickly in distilled water and air dry at room temperature.*
9. *Dip slide in Xylene or Xylene Substitute.*
10. *Mount in synthetic resin.*

References:

1. *Sheehan, D., Hrapchak, B., Theory and Practice of Histotechnology: 2nd Edition, 1980, pages 155-156.*
2. *A.F.I.P. Laboratory Methods in Histotechnology; 1992, pages 111.*
3. *Laboratory Medicine: Vol. 25, No. 6, June 1994, page 389.*

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