

Instructions For Use LBC-2-IFU

Rev. Date: July 17, 2011

Revision: 1

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P.O. Box 3286 - Logan, Utah 84323, U.S.A. - Tel. (800) 729-8350 - Tel. (435) 755-9848 - Fax (435) 755-0015 - www.scytek.com

Luxol Fast Blue Stain Kit

Description: The Luxol Fast Blue Stain Kit is designed for staining myelin/myelinated axons and Nissil substance on

formalin fixed, paraffin-embedded tissue as well as frozen tissue. This product is used for identifying the

basic neuronal structure in brain or spinal cord sections.

Myelinated Fibers: Blue
Nissil Substance: Violet
Nerve Cells: Violet

Uses/Limitations: Not to be taken internally.

For In-Vitro Diagnostic use only.

Histological applications.

Do not use past expiration date. Use caution when handling reagents.

Non-Sterile.

Control Tissue: Cerebral Cortex

Spinal Cord

Availability/Contents:

Item #	Kit Contents	<u>Volume</u>	Storage
CEA030	Cresyl Echt Violet Solution	30 ml	2-8℃.
LFB060	Luxol Fast Blue Solution	60 ml	18-25℃.
LCQ060	Lithium Carbonate Solution (0.05%)	60 ml	18-25℃.
EAS060	Alcohol, Reagent (70%)	60 ml	18-25℃.

Precautions: Avoid contact with skin and eyes.

May cause burns. Harmful if swallowed.

Follow all Federal, State, and local regulations regarding disposal.

Use in chemical fume hood whenever possible.

Procedure (Standard):

- 1. Deparaffinize sections if necessary and hydrate to distilled water.
- 2. Prepare humidity chamber by placing filter paper or other moisture absorbent paper towel in petri dish or other appropriate container. Thoroughly wet absorbent paper with DI/Distilled water and place slide on top of wet paper.
- 3. Apply 8-10 drops of Luxol Fast Blue Solution to tissue section, cover chamber with lid and incubate for a minimum of 2 hours at 60 ℃. Note: May be left at 60 ℃ overnight.
- 4. Remove slide from incubation chamber. If Luxol Fast Blue Solution has dried, rinse with 95% Ethanol to remove crystals.
- 5. Rinse thoroughly in distilled water.
- 6. Differentiate section by applying continuous drops of Lithium Carbonate Solution (0.05%) for up to 20 seconds.
- 7. Continue differentiation by continuously applying drops of Alcohol, Reagent (70%) until gray-matter is colorless and white-matter remains blue.
- 8. Rinse slide in distilled water.

Storage: 2° C

Mixed Storage Conditions.

Separate Contents.

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- 9. Incubate slide in 4-5 drops of Cresyl Echt Violet (0.1%) for 2-5 minutes.
- 10. Rinse quickly in 1 change of distilled water.
- 11. Dehydrate quickly in 3 changes of absolute alcohol.
- 12. Clear as desired and mount in synthetic resin.

References:

- 1. Sheenan, D.C., Hrapchak, B.B. Theory and Practice of Histotechnology, 2nd Edition. Battelle Press, Columbus, OH. Page 262-264. 1980
- 2. Kluver, H., Barrera, E.A. A Method for the combined staining of cells and fibers in the nervous system. Journal of Neuropathology and Experimental Neurology, 1953, 12: pages 400-403.

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