P.O. Box 3286 - Logan, Utah 84323, U.S.A. - Tel. (800) 729-8350 - Fax (435) 755-0015 - www.scytek.com

Phosphate Buffer Solution, pH 6.8
(For Giemsa Stain Kit)

Description: Phosphate Buffer Solution, pH 6.8 is a component of the Giemsa Stain Kit (Catalog# GMG-1) and is intended for use in the visualization of cells present in hematopoietic tissues and certain microorganisms. This kit may be used on formalin-fixed, paraffin-embedded or frozen sections.

Nuclei:	Blue/Violet
Cytoplasm	Light Blue
Collagen:	Pale Pink
Muscle Fibers:	Pale Pink
Erythrocytes:	Gray, Yellow or Pink
Rickettsia:	Reddish-Purple
Helicobacter Pylori:	Blue
Mast Cells:	Dark Blue with Red Granules

Uses/Limitations:
For In-Vitro Diagnostic use only.
Histological applications.
Do not use past expiration date.
Use caution when handling these reagents.

Control Tissue:
Blood Film
Any well fixed tissue.

Availability/Contents:

Item #	Kit Contents	Volume	Storage
PBM500	Phosphate Buffer Solution, pH 6.8	500 ml	Room Temperature
PBM999	Phosphate Buffer Solution, pH 6.8	1000 ml	Room Temperature

Required But Not Included:

MAY500	May-Grunwald Stock Solution	500 ml	Room Temperature
GGS500	Giemsa Stock Solution	500 ml	Room Temperature

Precautions:
Keep away from open flame.
Avoid contact with skin and eyes.
Harmful if swallowed.
Follow all Federal, State, and local regulations regarding disposal.
Use in chemical fume hood whenever possible.

Storage: 18° C  25° C

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Preparation of Reagents Prior to Beginning:

1. Prepare **Working May-Grunwald Solution** by mixing 25ml of May-Grunwald Solution (MAY500) with 25ml of Phosphate Buffer Solution, pH 6.8 (PBM500).

2. Prepare **Working Giemsa Solution** by mixing 2.5ml of Giemsa Stock Solution (GGS500) with 50ml of Phosphate Buffer Solution, pH 6.8 (PBM500).

Procedure (Standard):

1. Deparaffinize sections if necessary and hydrate to distilled water.
2. Place slide in staining tray and flood with Working May-Grunwald Solution for 6 minutes. Note: Agitate slide occasionally to insure proper staining.
3. Flood slide with Phosphate Buffer Solution, pH 6.8 until no stain runs off.
4. Flood slide with Working Giemsa Solution for 13 minutes. Note: Agitate slide occasionally to insure proper staining.
5. Flood slide with Phosphate Buffer Solution, pH 6.8 until no stain runs off.
6. Allow slide to remain in Phosphate Buffer Solution, pH 6.8 for an additional 3 minutes.
7. Dip slide quickly in distilled water and air dry at room temperature.
8. Dip slide in Xylene or Xylene Substitute.
9. Mount in synthetic resin.

Procedure (Mast Cells):

1. Deparaffinize sections if necessary and hydrate to distilled water.
2. Place slide in staining tray and flood with Working May-Grunwald Solution for 6 minutes. Note: Agitate slide occasionally to insure proper staining.
3. Flood slide with Phosphate Buffer Solution, pH 6.8 until no stain runs off.
4. Flood slide with Working Giemsa Solution for 13 minutes. Note: Agitate slide occasionally to insure proper staining.
5. Flood slide with Phosphate Buffer Solution, pH 6.8 until no stain runs off.
6. Differentiate by dipping slide in Acetic Acid Solution (0.25%) until background is desired intensity.

Storage: 18° C  25° C

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7. Dip slide 20 times in Phosphate Buffer Solution, pH 6.8.
8. Dip slide quickly in distilled water and air dry at room temperature.
9. Dip slide in Xylene or Xylene Substitute.
10. Mount in synthetic resin.

References:

1. Sheehan, D., Hrapchak, B., Theory and Practice of Histotechnology: 2nd Edition, 1980, pages 155-156.
2. A.F.I.P. Laboratory Methods in Histotechnology; 1992, pages 111.
3. Laboratory Medicine: Vol. 25, No. 6, June 1994, page 389.