



Cod. CM140

REV:16-06-2020

### Technical-application suggestions

#### Needle-Aspirated Samples

Synthetically the preparation process of the material coming from the needle-aspirated is the follow:

1. Withdrawal with needle aspiration from the structure of interest.
1. Deposition of the aspirated material on CytoMatrix (1-2 drops ).
1. Open the biocassette in which CytoMatrix is contained
2. \*Deposit the drawn needle material on the surface bounded by the frame
2. Close the biocassette and immerse it in formalin for at least 12 hours
3. The CytoMatrix-needle aspiration material complex is processed and paraffin embedded as any tissue sample (long cycle processing)
4. **In the inclusion phase remember to orient the part with the "Frame" towards the bottom of the tray**
5. Apply to the inclusion obtained, the various diagnostic techniques used in the histopathology laboratory

(\*): The deposition of the sample on the CytoMatrix could lead to the formation of a small drop which could reduce the absorption of the sample by the matrix. It is suggested to act with the tip of the needle on the drop itself trying to lengthen it in some direction. This maneuver will lead to the drop breaking with quick absorption of the sample (1 minute). The presence of microfrustles in the sampling will cause these to be placed on the surface on the matrix retained by its positive charge. The immersion in formalin will lead to the formation of chemical bonds between the sample and the matrix which will make the complex a whole. In this case, being the sample tied on the surface, it is suggested to collect the material on the slides as soon as it appears at the cut, without waiting to reach the end of the frame.

(\*\*): The collected material may not be abundant, so it is advisable to pay particular attention to the first sections obtained with CytoMatrix.

It is recommended to cut 10 sections in sequence. Load sections 1 and 10 on the first slide and stain it with EE. The intermediate sections will be loaded individually on the others 8 slides. The presence of sample on both sections of the first slide, will confirm the presence of sample in the others 8 slides

**Use positively charged slides for better adhesion of the sections to the glass**

### Technical-application suggestions

#### Effusions, Urine,

Synthetically, the process of preparing the sample from an effusion (pleural abdominal, pericardial) or urine requires:

1. (#) Place the sample in a 50ml "Falcon" and centrifuge at 1600 RFM for 10 minutes
2. Remove the supernatant and add the fixative used in the lab to the "Cell Pellet" obtained in a 1:1 ratio volume (a mixture of 100° alcohol/buffered :formalin in 1:1 ratio is recommended as fixative)
3. Centrifuge at 1600 rpm per 10 minuti
4. Remove the supernatant
5. \*Deposit 50ul of "cell pellet " on CytoMatrix
6. Close the biocassette and immerse it in formalin for at least 6-8 hours .
7. The CytoMatrix-sample complex is processed and paraffin embedded as any tissue sample.
8. **During the inclusion phase remember to orient the part with the "Frame" towards the bottom of the tray**
9. Apply, the various diagnostic techniques used in the histopathology laboratory to the inclusion obtained .

(#): In the event of a blood effusion it is suggested to add lysing to the sample in 1:1 volumetric ratio.

(\*): The deposition of the sample on the CytoMatrix could lead to the formation of a small drop which could reduce the absorption of the sample by the matrix. It is suggested to act with the tip of the needle on the drop itself trying to lengthen it in some direction. This maneuver will lead to the drop breaking with quick absorption of the sample (1 minute).

(\*\*):As the collected material may not be abundant, it is advisable to pay particular attention to the first sections obtained with CytoMatrix.

It is recommended to cut more sections in sequence. Load sections 1 and last on the first slide and stain it with EE. The intermediate sections will be loaded individually on the other slides. The presence of sample on both sections of the first slide, will confirm the presence of sample in the other slides

**Use positively charged slides for better adhesion of the sections to the glass**

CE-IVD

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UCS Diagnostic Srl & University Campus Biomedico of Rome

[www.cytomatrix.it](http://www.cytomatrix.it)